Effects of Oral Branched-Chain Amino Acids on Hepatic Encephalopathy and Outcome in Patients With Liver Cirrhosis

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Abstract

Branched-chain amino acids (BCAAs) constituting of valine, leucine, and isoleucine act as both substrates of proteins and as key regulators for various nutrient metabolisms. Patients with liver cirrhosis frequently lack sufficient BCAAs and therefore suffer from various metabolic disorders. Hepatic encephalopathy (HE) is a severe metabolic disorder with neurologic manifestations such as flapping tremors and coma in patients with liver cirrhosis. In addition, a mild form of HE known as minimal HE (MHE) is an important social issue because it occurs in up to 80% of patients with chronic liver disease and affects prognosis and activities of daily living, possibly resulting in falls and motor vehicle accidents. Although HE/MHE can be caused by various pathological conditions, including in an accumulation of mercaptans, short-chain fatty acids, and alterations in the gut flora, hyperammonemia has also been implicated in an important pathogenesis of HE/MHE. Besides urea cycle of liver, ammonia can be detoxified in the skeletal muscles by the amidation process for glutamine synthesis using BCAAs. Thus, BCAA supplementation may enhance detoxification of ammonia in skeletal muscle and may be a possible therapeutic strategy for HE/MHE. In this review, we summarize the clinical impacts of BCAA supplementation on HE/MHE and discuss possible mechanisms for a BCAA-induced improvement of HE/MHE. Furthermore, we present some modifications of oral BCAA therapy for improvement of efficacy in HE treatment. We also briefly describe pleiotropic benefits of BCAAs on lifethreatening events and overall prognosis in patients with liver cirrhosis. *(Nutr Clin Pract.* 2013;28:580-588)

Keywords

nutrition therapy; ammonia; hepatic encephalopathy; cognitive impairment; branched chain amino acids; liver diseases; end-stage liver disease

Introduction

The amino acid valine, leucine, and isoleucine are known as branched-chain amino acids (BCAAs) because of their structural features. They are the most abundant essential amino acids, comprising approximately 40%. BCAAs serve as substrates for the synthesis of body proteins similar to the other amino acids. In addition, BCAAs are known to regulate various nutrient metabolism pathways.¹⁻⁶

BCAAs have been suggested to be associated with the development of hepatic encephalopathy through alteration in ammonia and energy metabolisms. BCAAs are first catabolized, mostly in skeletal muscles, via BCAA aminotransferase (BCAT) with α -ketoglutarate into glutamate and branchedchain keto acids (BCKAs).⁷⁻⁹ Although glutamate forms alanine in the presence of pyruvate and alanine aminotransferase, it also forms glutamine via glutamine synthetase (GS) and detoxifies ammonia when there are increased levels of ammonia (Figure 1), contributing to the improvement of hyperammonemia and hepatic encephalopathy.^{$7-9$} The other metabolites of the reaction, BCKAs, then undergo oxidative decarboxylation, catalyzed by BCKA dehydrogenase (BCKDH), to form the corresponding branched-chain acylcoenzyme A (acyl-CoA) esters.7-9 Then, branched-chain acyl-CoA is metabolized in the tricarboxylic acid (TCA) cycle, resulting in energy production. The energy production occurs in neurons and astrocytes, contributing to the improvement of cerebral activity and hepatic encephalopathy (Figure 1).⁷⁻⁹

Recently, BCAAs have been investigated as pharmacological agents.^{1,3-6} BCAAs, particularly Leu, activate the mammalian target of rapamycin and subsequently upregulates the downstream eukaryotic initiation factor 4E-binding protein-1 and 70-kDa ribosomal protein S6 kinase, which regulate mRNA translation and synthesis of protein.^{1,10-12} BCAAs also

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Figure 1. The metabolism of BCAAs in the skeletal muscle. BCAAs, branched-chain amino acids; BCAT, BCAA aminotransferase; NH_4^+ , ammonia; ATP, adenosine 5'-triphosphate; ADP, adenosine 5'-diphosphate; BCKAs, branched-chain keto acids; BCKDH, BCKA dehydrogenase; Acyl-CoA, acylcoenzyme A esters; TCA, tricarboxylic acid.

upregulate both intracellular insulin signaling molecules including phosphatidylinositol 3-kinase and liver X receptor α / sterol regulatory element binding protein-1c pathway, resulting in improvement of glucose metabolism/insulin resistance.^{1,13-17} They also control food intake or energy balance via action on central nervous system.¹⁸⁻²⁰ In particular, on the protein metabolism, leucine among the BCAAs seems to play an important role in the protein synthesis in several tissues including the liver, skeletal muscle, and adipose tissue.²¹⁻²³ Furthermore, leucine and isoleucine seem to be associated with glucose metabolism in the liver and skeletal muscle.^{$24-26$} Thus, BCAAs exert various effects on nutrient metabolisms.

Metabolic disturbances in carbohydrates, lipids, or amino acids and proteins pathways, such as hepatic encephalopathy (HE), are evident in patients with liver disease.²⁷⁻²⁹ Because the serum level of BCAAs, along with BCAA-to-aromatic amino acid (AAA) ratio, decreases in these patients, supplementation of BCAAs is an attractive potential treatment strategy to improve these metabolic disturbances. In this review, we describe the impact of BCAAs on the prevention and treatment of HE. In addition, we briefly summarize pleiotropic benefits of BCAAs for cirrhotic patients.

Diagnosis of HE

HE is a common complication of fulminant hepatitis or cirrhosis, characterized by various neurological manifestations. Therefore, HE is generally diagnosed by the presence of a depressed level of consciousness and neurological abnormalities after the exclusion of the other brain diseases. Recently, a new nomenclature of HE was proposed on the basis of the type of hepatic abnormality and the duration or characteristics of neurologic manifestations.30 Three types of hepatic abnormalities are associated with HE.³⁰ Episodic HE is characterized by

the HE between the HE episodes, whereas persistent HE is characterized by the lack of quiescent HE. Minimal HE (MHE) is defined as HE remaining below the clinically detectable levels.³¹⁻³³

The severity of HE is subjectively assessed because of the absence of objective criteria. The West-Haven criteria are most widely used for grading the severity of HE.³⁰ Grades 2 to 4 of HE can be simply and clinically diagnosed because the presence of neurologic manifestations is overt, whereas grade 1 of HE is usually diagnosed retrospectively because no specific signs and symptoms pertaining specifically to grade 1 of HE exist. Therefore, it is clinically difficult to distinguish grade 1 of HE from a normal state (grade 0). Furthermore, MHE could be latent in grade 0 because MHE is defined as cognitive dysfunction without clinical signs of overt HE. Hence, clinical distinction among normal, MHE, and grade 1 of HE is difficult.

MHE is usually diagnosed by neuropsychological tests with at least 2 of the following neuropsychological tests: 30° a battery using number connection test (NCT) A and B, blockdesign test, and digit-symbol test or a standardized test battery including NCT A and B, line-tracing test, serial-dotting test, and digitsymbol test (PSE-Syndrome-Test). Although equipment is required, quantitative neurophysiological tools such as electroencephalography with mean dominant frequency, P300 auditory evoked potentials are also recommended.^{30,34-40}

Effects of Oral BCAAs on HE

HE can be caused by various pathological conditions, including dehydration, spontaneous bacterial peritonitis, noncompliance of medication, an accumulation of mercaptans, short-chain fatty acids, γ-aminobutyric acid, benzodiazepines, and alterations in the gut flora, blood brain barrier, and regional differences in cerebral glucose metabolism.⁴¹⁻⁵⁰ In addition, a decrease in serum BCAA levels has been implicated in one of the pathogenesis of HE in cirrhotic patients.^{5,51-53} Although lactulose or antibiotics are established treatment for HE, BCAA supplementation seems to further ameliorate HE in cirrhotic patients.

Although BCAAs are currently prescribed as an oral preparation,⁵⁴ the effects of oral BCAA supplementation on HE remain controversial, as summarized in Table 1. Four randomized controlled trials demonstrated that oral BCAA supplementation does not have beneficial effects on HE;⁵⁵⁻⁵⁷ however, the number of enrolled patients in the BCAA arm of these studies is small $(n < 10)$, and therefore these studies did not detect a difference between BCAA-treated and control groups. In fact, Ichida et al performed a multicenter study, enrolling a large number of cirrhotic patients ($n = 96$) to evaluate the efficacy of long-term administration of oral BCAA supplementation.58 The study demonstrated that BCAA supplementation significantly increases BCAA-to-AAA ratio and decreases the hepatic encephalopathic grade.⁵⁸ In addition, Horst et al conducted a randomized study comparing dietary protein with an

Reference			Dose of BCAAs or		
	n	Control	Control	Trial period	Improvement of HE
Sieg et al 57	14	Carbohydrate	13.2 g/day	3 months	No
Horst et al^{59}	37	Dietary protein	$20-60$ g/day	4 weeks	Yes
Marchesini et al ⁶⁰	34	Casein	0.24 g/kg/day	3 months	Yes
Bianchi et al ⁶¹	49	Casein	0.24 g/kg/day	3 months	Yes

Table 1. Randomized Control Trials of Oral BCAA Supplementation on HE in Cirrhotic Patients.

BCAAs, branched-chain amino acids; HE, hepatic encephalopathy.

Table 2. Randomized Control Trials of Oral BCAA Supplementation on MHE in Cirrhotic Patients.

Reference		Control	Dose of BCAAs	Trial Period	Improvement of MHE
Plauth et $al63$	¹⁶	No amino acids	$3 \text{ g}/day$	8 weeks	Yes
Les et al 64		Maltodextrin	$30 \frac{g}{day}/day$	56 weeks	Yes

BCAAs, branched-chain amino acids; MHE, minimal hepatic encephalopathy.

oral BCAA supplement and showed a significant improvement in the mental status grade, flapping tremors, and portal-systemic encephalopathy index.⁵⁹ Similarly, 2 randomized double-blind casein-controlled trials conducted by Marchesini et al and Bianchi et al demonstrated that oral BCAA supplementation improves both nutrition parameters and the mental state of cirrhotic patients with chronic encephalopathy.^{60,61} Therefore, oral administration of BCAAs may have beneficial effects on HE.

Effects of BCAAs on MHE

MHE, which is known as a subclinical or latent HE, is considered as the mildest form of the clinical spectrum of HE.^{31,32} Kato et al demonstrated that nutrition consultations for energy and protein intakes improve MHE, and the severity of MHE is negatively associated with serum BCAA levels.⁶² Thus, BCAA supplementation is considered as a therapeutic agent for MHE. In fact, 2 randomized controlled trials conducted Egberts et al and Plauth et al examined the effects of oral BCAA supplementation on MHE (Table 2) and showed that BCAA supplementation improves psychomotor functions, attention, practical intelligence, and automobile driving capacity. 63 Recently, Les et al conducted a randomized control trial to investigate the long-term effects of BCAA supplementation on the recurrence of HE^{64} (Table 2). Although BCAA does not decrease the recurrence rate of HE in the study, BCAA improves MHE, evaluated by Trail Making Test part A, Digit Symbol Test, and Grooved Pegboard Test.⁶⁴ Thus, all of these randomized control trials corroborated the beneficial effects of BCAA supplementation in MHE. Since MHE occurs in up to 80% of patients with chronic liver disease⁶⁵ and affects daily living, possibly leading to falls, motor vehicle accidents, and prognosis, 38,39,65 BCAA supplementation may improve both MHE and the quality of life or prognosis in a great number of cirrhotic patients.

Mechanisms for BCAA-Caused Improvement of HE

Although the precise mechanisms for BCAA-caused improvement of HE remains unknown, several potential mechanisms are proposed (Figure 2). First, BCAAs enhance detoxification of blood ammonia in skeletal muscles.⁶⁶ In cirrhotic patients, increased blood ammonia is detoxified both in the liver and in the skeletal muscles, which is caused by liver dysfunction. 67 In skeletal muscles, BCAAs are catabolized to glutamate, which incorporates ammonia in the amidation process for glutamine synthesis, leading to a decrease in blood ammonia levels.68 Thus, the skeletal muscle stimulatory effect and its potential role in ameliorating HE is being increasingly recognized. Second, BCAAs improve cerebral hyperammonemia.⁶⁹ Cerebral hyperammonemia causes impairment of neurotransmission and depletion of cerebral energy production as well as astrocyte swelling, leading to $HE⁷⁰$ In both neurons and astrocytes, isoleucine is metabolized to acetyl-CoA and succinyl-CoA, which produce adenosine triphosphate (ATP) by enhancing TCA cycle metabolism and is subsequently metabolized to glutamate and aspartate. These amino acids serve as anaplerotic substances for the detoxification of cerebral ammonia. Third, BCAAs reduce brain uptake of AAAs and serotonin-precursor tryptophan. AAAs alter intracerebral synthesis of serotonin and produce octopamine and phenylethylamine, which act as false neurotransmitters, leading to $HE⁷¹BCAAs$ compete with AAAs and tryptophan for the amino acid transporter across the blood-brain barrier.⁷² In addition to these favorable effects, BCAAs are known to promote liver

Figure 2. Mechanisms for BCAA-caused improvement of HE. BCAAs, branched-chain amino acids; ATP, adenosine 5'-triphosphate; TCA, tricarboxylic acid.

regeneration^{73,74} and improve cerebral perfusion in patients with cirrhosis.⁷⁵⁻⁷⁶ These effects may also be involved in the BCAA-induced improvement of HE.

Additive Effects of Zinc and Amino Acids to Oral BCAA Supplementation on HE

Zinc is a coenzyme of ornithine transcarbamylase (OTC), which is a key enzyme in the urea cycle of the liver.⁷⁷ Since cirrhotic patients are deficient in zinc, OTC activity is decreased, resulting in an increase of blood ammonia levels through the down-regulation of urea cycle.^{78,79} In cirrhotic rats, oral zinc supplementation increased hepatic OTC activity, leading to a decrease in blood ammonia levels.^{80,81} In cirrhotic patients, BCAAs in combination with zinc administration significantly decreased blood ammonia levels and improved HE compared to BCAA supplementation alone. $82,83$

The additive effects of other amino acids to oral BCAA supplementation have recently been investigated. Ndraha et al performed a randomized controlled, double-blinded study and demonstrated that the addition of L -ornithine L -aspartate (LOLA) to BCAA supplementation enhances the improvement of HE.84 A possible mechanism is that LOLA stimulates the urea cycle and glutamine synthesis, which enhances the detoxification of ammonia.⁸⁵ Malaguarnera et al showed that the addition of _L-acetylcarnitine to BCAA supplementation significantly improves HE and decreases blood ammonia levels in a randomized controlled double-blind study.⁸⁶

Other Favorable Effects of BCAAs

BCAA supplementation was originally invented in order to improve the imbalance of amino acids and nutrition status. However, recent studies have revealed that BCAAs modulate

Effects	Number of Studies	References
Enhancement of albumin synthesis	19	4,16,90-103
Suppression of the development of HCC	16	104-119
Improvement of glucose metabolism/insulin resistance	12	16,17,107,120-128
Enhancement of immunity	7	129-135
Enhancement of liver regeneration	6	73,136-140
Suppression of tumor-associated angiogenesis	4	107,111,115,141
Suppression of hepatic fibrosis	3	90,142,143
Suppression of oxidative stress	\overline{c}	144,145
Suppression of HCV replication	2	146,147
Improvement of QOL	5	4,74,148-152

Table 3. Pleiotropic Effects of BCAAs.

BCAA, branched-chain amino acids; HCC, hepatocellular carcinoma; HCV, hepatitis C virus; QOL, quality of life.

various intracellular signaling including mammalian target of rapamycin and insulin signaling pathways and exert diverse beneficial effects for cirrhotic patients.^{1,2,5,9,53,87,88}

As shown in Table 3, many studies indicated that BCAAs demonstrate favorable effects for albumin synthesis, hepatocarcinogenesis, glucose intolerance/insulin resistance, immunity, and liver regeneration. Early evidence also suggests that BCAAs suppress angiogenesis, hepatic fibrosis, oxidative stress, and hepatitis C virus replication. Thus, we hypothesize that BCAAs may prolong the overall survival of cirrhotic patients. Marchesini et al and Muto et al demonstrated that BCAAs prevented progressive hepatic failure and improved event-free survival in cirrhotic patients, respectively.^{3,4} Moreover, a Japanese nationwide study recently revealed that BCAAs prolonged survival of cirrhotic patients by suppressing the onset of life-threatening events.⁸⁹ The study also disclosed that in addition to serum alpha-fetoprotein levels and amino acid imbalance, BCAA administration is identified as an independent negative risk factor for life-threatening events in cirrhotic patients.⁸⁹

Conclusion

In this review, we summarized the clinical impacts of BCAA supplementation on HE/MHE and discussed possible mechanisms for BCAA-induced improvement of HE/MHE. The mainstay therapy for HE and MHE is lactulose and antibiotics. BCAAs are expensive and the clinical significance was limited because of small numbers of patients. Thus, BCAA may be an adjunct or alternative to the mainstay therapy for HE and MHE. However, BCAAs serve other beneficial effects on albumin synthesis and development of hepatocellular carcinoma in

cirrhotic patients. BCAAs also have potential to improve hepatic fibrosis, insulin resistance, immunity, and patients' quality of life, suggesting that BCAAs seem to have good influence on the management of cirrhotic patients.

References

- 1. Kawaguchi T, Izumi N, Charlton MR, Sata M. Branched-chain amino acids as pharmacological nutrients in chronic liver disease. *Hepatology*. 2011;54(3):1063-1070.
- 2. Charlton M. Branched-chain amino acid enriched supplements as therapy for liver disease. *J Nutr*. 2006;136(suppl 1):295S-298S.
- 3. Marchesini G, Bianchi G, Merli M, et al. Nutritional supplementation with branched-chain amino acids in advanced cirrhosis: a double-blind, randomized trial. *Gastroenterology*. 2003;124(7):1792-1801.
- 4. Muto Y, Sato S, Watanabe A, et al. Effects of oral branched-chain amino acid granules on event-free survival in patients with liver cirrhosis. *Clin Gastroenterol Hepatol*. 2005;3(7):705-713.
- 5. Plauth M, Schutz T. Branched-chain amino acids in liver disease: new aspects of long known phenomena. *Curr Opin Clin Nutr Metab Care*. 2011;14(1):61-66.
- 6. Yoshizawa F. New therapeutic strategy for amino acid medicine: notable functions of branched chain amino acids as biological regulators. *J Pharmacol Sci*. 2012;118(2):149-155.
- 7. Paul HS, Adibi SA. Activation of hepatic branched chain alphaketo acid dehydrogenase by a skeletal muscle factor. *J Biol Chem*. 1982;257(21):12581-12588.
- 8. Damuni Z, Merryfield ML, Humphreys JS, Reed LJ. Purification and properties of branched-chain alpha-keto acid dehydrogenase phosphatase from bovine kidney. *Proc Natl Acad Sci U S A*. 1984;81(14):4335- 4338.
- 9. Holecek M. Three targets of branched-chain amino acid supplementation in the treatment of liver disease. *Nutrition*. 2010;26(5):482-490.
- 10. Tischler ME, Desautels M, Goldberg AL. Does leucine, leucyl-tRNA, or some metabolite of leucine regulate protein synthesis and degradation in skeletal and cardiac muscle? *J Biol Chem*. 1982;257(4):1613-1621.
- 11. Kimball SR, Jefferson LS. Regulation of protein synthesis by branchedchain amino acids. *Curr Opin Clin Nutr Metab Care*. 2001;4(1):39-43.
- 12. Yoshizawa F. Regulation of protein synthesis by branched-chain amino acids in vivo. *Biochem Biophys Res Commun*. 2004;313(2):417-422.
- 13. Sener A, Malaisse WJ. The stimulus-secretion coupling of amino acidinduced insulin release: insulinotropic action of branched-chain amino acids at physiological concentrations of glucose and glutamine. *Eur J Clin Invest*. 1981;11(6):455-460.
- 14. Tappy L, Acheson K, Normand S, et al. Effects of infused amino acids on glucose production and utilization in healthy human subjects. *Am J Physiol*. 1992;262(6 Pt 1):E826-E33.
- 15. Nishitani S, Takehana K, Fujitani S, Sonaka I. Branched-chain amino acids improve glucose metabolism in rats with liver cirrhosis. *Am J Physiol Gastrointest Liver Physiol*. 2005;288(6):G1292-G1300.
- 16. Kawaguchi T, Nagao Y, Matsuoka H, Ide T, Sata M. Branched-chain amino acid-enriched supplementation improves insulin resistance in patients with chronic liver disease. *Int J Mol Med*. 2008;22(1):105-112.
- 17. Takeshita Y, Takamura T, Kita Y, et al. Beneficial effect of branchedchain amino acid supplementation on glycemic control in chronic hepatitis C patients with insulin resistance: implications for type 2 diabetes. *Metabolism*. 2012;61(10):1388-1394.
- 18. Cota D, Proulx K, Smith KA, et al. Hypothalamic mTOR signaling regulates food intake. *Science*. 2006;312(5775):927-930.
- 19. She P, Reid TM, Bronson SK, et al. Disruption of BCATm in mice leads to increased energy expenditure associated with the activation of a futile protein turnover cycle. *Cell Metab*. 2007;6(3):181-194.
- 20. Su Y, Lam TK, He W, et al. Hypothalamic leucine metabolism regulates liver glucose production. *Diabetes*. 2012;61(1):85-93.
- 21. Anthony JC, Yoshizawa F, Anthony TG, et al. Leucine stimulates translation initiation in skeletal muscle of postabsorptive rats via a rapamycinsensitive pathway. *J Nutr*. 2000;130(10):2413-2419.
- 22. Anthony TG, Anthony JC, Yoshizawa F, Kimball SR, Jefferson LS. Oral administration of leucine stimulates ribosomal protein mRNA translation but not global rates of protein synthesis in the liver of rats. *J Nutr*. 2001;131(4):1171-1176.
- 23. Lynch CJ, Patson BJ, Anthony J, et al. Leucine is a direct-acting nutrient signal that regulates protein synthesis in adipose tissue. *Am J Physiol Endocrinol Metab*. 2002;283(3):E503-E513.
- 24. Doi M, Yamaoka I, Nakayama M, et al. Isoleucine, a blood glucoselowering amino acid, increases glucose uptake in rat skeletal muscle in the absence of increases in AMP-activated protein kinase activity. *J Nutr*. 2005;135(9):2103-2108.
- 25. Doi M, Yamaoka I, Nakayama M, Sugahara K, Yoshizawa F. Hypoglycemic effect of isoleucine involves increased muscle glucose uptake and whole body glucose oxidation and decreased hepatic gluconeogenesis. *Am J Physiol Endocrinol Metab*. 2007;292(6):E1683-E1693.
- 26. Peyrollier K, Hajduch E, Blair AS, Hyde R, Hundal HS. L-leucine availability regulates phosphatidylinositol 3-kinase, p70 S6 kinase and glycogen synthase kinase-3 activity in L6 muscle cells: evidence for the involvement of the mammalian target of rapamycin (mTOR) pathway in the L-leucine-induced up-regulation of system A amino acid transport. *Biochem J*. 2000;350(Pt 2):361-368.
- 27. Lam VW, Poon RT. Role of branched-chain amino acids in management of cirrhosis and hepatocellular carcinoma. *Hepatol Res*. 2008;38(s1The 6 Japan Society of Hepatology Single Topic Conference: Liver Failure: Recent Progress and Pathogenesis to Management. 28-29 September 2007, Iwate, Japan):S107-S115.
- 28. Kawaguchi T, Yoshida T, Harada M, et al. Hepatitis C virus down-regulates insulin receptor substrates 1 and 2 through up-regulation of suppressor of cytokine signaling 3. *Am J Pathol*. 2004;165(5):1499-1508.
- 29. Kawaguchi T, Ide T, Taniguchi E, et al. Clearance of HCV improves insulin resistance, beta-cell function, and hepatic expression of insulin receptor substrate 1 and 2. *Am J Gastroenterol*. 2007;102(3):570-576.
- 30. Ferenci P, Lockwood A, Mullen K, et al. Hepatic encephalopathy—definition, nomenclature, diagnosis, and quantification: final report of the working party at the 11th World Congresses of Gastroenterology, Vienna, 1998. *Hepatology*. 2002;35(3):716-721.
- 31. Dhiman RK, Saraswat VA, Sharma BK, et al. Minimal hepatic encephalopathy: consensus statement of a working party of the Indian National Association for Study of the Liver. *J Gastroenterol Hepatol*. 2010;25(6):1029-1041.
- 32. Bajaj JS, Cordoba J, Mullen KD, et al. Review article: the design of clinical trials in hepatic encephalopathy—an International Society for Hepatic Encephalopathy and Nitrogen Metabolism (ISHEN) consensus statement. *Aliment Pharmacol Ther*. 2011;33(7):739-747.
- 33. Kato A, Watanabe Y, Sawara K, Suzuki K. Diagnosis of sub-clinical hepatic encephalopathy by Neuropsychological Tests (NP-tests). *Hepatol Res*. 2008;38(suppl 1):S122-S127.
- 34. Sawara K, Desjardins P, Chatauret N, et al. Alterations in expression of genes coding for proteins of the neurovascular unit in ischemic liver failure. *Neurochem Int*. 2009;55(1-3):119-123.
- 35. Kato A, Suzuki K, Kaneta H, et al. Regional differences in cerebral glucose metabolism in cirrhotic patients with subclinical hepatic encephalopathy using positron emission tomography. *Hepatol Res*. 2000;17(3):237-245.
- 36. Ciecko-Michalska I, Wojcik J, Wyczesany M, et al. Cognitive evoked response potentials in patients with liver cirrhosis without diagnosis of minimal or overt hepatic encephalopathy. A pilot study. *J Physiol Pharmacol*. 2012;63(3):271-276.
- 37. Seo YS, Yim SY, Jung JY, et al. The Psychometric Hepatic Encephalopathy Score for the detection of minimal hepatic encephalopathy in Korean patients with liver cirrhosis. *J Gastroenterol Hepatol*. 2012;27(11):1695- 1704.
- 38. Kawaguchi T, Taniguchi E, Sata M. Motor vehicle accidents: how should cirrhotic patients be managed? *World J Gastroenterol*. 2012;18(21):2597- 2599.
- 39. Bajaj JS, Pinkerton SD, Sanyal AJ, Heuman DM. Diagnosis and treatment of minimal hepatic encephalopathy to prevent motor vehicle accidents: a cost-effectiveness analysis. *Hepatology*. 2012;55(4):1164-1171.
- 40. Taniguchi E, Kawaguchi T, Sakata M, et al. Lipid profile is associated with the incidence of cognitive dysfunction in viral cirrhotic patients: a data-mining analysis. *Hepatol Res*. 2013;43(4):418-424.
- 41. Ferenci P, Ebner J, Zimmermann C, et al. Overestimation of serum concentrations of gamma-aminobutyric acid in patients with hepatic encephalopathy by the gamma-aminobutyric acid-radioreceptor assay. *Hepatology*. 1988;8(1):69-72.
- 42. Clausen MR, Mortensen PB, Bendtsen F. Serum levels of short-chain fatty acids in cirrhosis and hepatic coma. *Hepatology*. 1991;14(6):1040-1045.
- 43. Basile AS, Jones EA. Ammonia and GABA-ergic neurotransmission: interrelated factors in the pathogenesis of hepatic encephalopathy. *Hepatology*. 1997;25(6):1303-1305.
- 44. Riordan SM, Williams R. Treatment of hepatic encephalopathy. *N Engl J Med*. 1997;337(7):473-479.
- 45. Matsuda Y, Fujita T, Hada T, Higashino K. Comparative study on the correlation of plasma gamma-aminobutyric acid and pipecolic acid with liver function in patients with liver cirrhosis. *Hepatol Res*. 2000;18(2):132-140.
- 46. Katayama K. Ammonia metabolism and hepatic encephalopathy. *Hepatol Res*. 2004;30(S):73-80.
- 47. Lighthouse J, Naito Y, Helmy A, et al. Endotoxinemia and benzodiazepine-like substances in compensated cirrhotic patients: a randomized study comparing the effect of rifaximine alone and in association with a symbiotic preparation. *Hepatol Res*. 2004;28(3):155-160.
- 48. Sato S, Tateishi K, Kato A, et al. Marked depression of brain cholecystokinin and vasoactive intestinal polypeptide levels in Eck fistula dogs. *Regul Pept*. 1989;25(1):111-121.
- 49. Tateishi K, Sato S, Kato A, et al. Reduced somatostatin-like immunoreactivity in the brain of dogs with an Eck fistula. *Regul Pept*. 1987;18(5- 6):277-286.
- 50. Tateishi K, Sato S, Kato A, et al. Reduced somatostatin-28-(1-12)-like immunoreactivity in cerebral cortex of dogs with an Eck fistula and somatostatin molecular forms in brain. *Regul Pept*. 1988;23(3):247-259.
- 51. Hayashi S, Watanabe A, Shiota T, et al. Effects of intraduodenal feeding of a branched-chain amino acid-rich solution on ammonia-induced encephalopathy in liver-injured rats. *Gastroenterol Jpn*. 1982;17(6):538- 541.
- 52. Suzuki K, Kato A, Iwai M. Branched-chain amino acid treatment in patients with liver cirrhosis. *Hepatol Res*. 2004;30(S):25-29.
- 53. Chadalavada R, Sappati Biyyani RS, Maxwell J, Mullen K. Nutrition in hepatic encephalopathy. *Nutr Clin Pract*. 2010;25(3):257-264.
- 54. Kato A, Suzuki K. How to select BCAA preparations. *Hepatol Res*. 2004;30:30-35.
- 55. Schafer K, Winther MB, Ukida M, et al. Influence of an orally administered protein mixture enriched in branched chain amino acids on the chronic hepatic encephalopathy (CHE) of patients with liver cirrhosis. *Z Gastroenterol*. 1981;19(7):356-362.
- 56. Eriksson LS, Persson A, Wahren J. Branched-chain amino acids in the treatment of chronic hepatic encephalopathy. *Gut*. 1982;23(10):801-806.
- 57. Sieg A, Walker S, Czygan P, et al. Branched-chain amino acid-enriched elemental diet in patients with cirrhosis of the liver. A double blind crossover trial. *Z Gastroenterol*. 1983;21(11):644-650.
- 58. Ichida T, Shibasaki K, Muto Y, et al. Clinical study of an enteral branchedchain amino acid solution in decompensated liver cirrhosis with hepatic encephalopathy. *Nutrition*. 1995;11(suppl 2):238-244.
- 59. Horst D, Grace ND, Conn HO, et al. Comparison of dietary protein with an oral, branched chain-enriched amino acid supplement in chronic portal-systemic encephalopathy: a randomized controlled trial. *Hepatology*. 1984;4(2):279-287.
- 60. Marchesini G, Dioguardi FS, Bianchi GP, et al. Long-term oral branchedchain amino acid treatment in chronic hepatic encephalopathy. A randomized double-blind casein-controlled trial. The Italian Multicenter Study Group. *J Hepatol*. 1990;11(1):92-101.
- 61. Bianchi GP, Marchesini G, Zoli M, et al. Oral BCAA supplementation in cirrhosis with chronic encephalopathy: effects on prolactin and estradiol levels. *Hepatogastroenterology*. 1992;39(5):443-446.
- 62. Kato A, Tanaka H, Kawaguchi T, et al. Nutritional Management contributes to improvement in minimal hepatic encephalopathy and quality of life in patients with liver cirrhosis; a preliminary, prospective open-label study. *Hepatol Res*. 2013;43(5):452-458.
- 63. Plauth M, Egberts EH, Hamster W, et al. Long-term treatment of latent portosystemic encephalopathy with branched-chain amino acids. A double-blind placebo-controlled crossover study. *J Hepatol*. 1993;17(3): 308-314.
- 64. Les I, Doval E, Garcia-Martinez R, et al. Effects of branched-chain amino acids supplementation in patients with cirrhosis and a previous episode of hepatic encephalopathy: a randomized study. *Am J Gastroenterol*. 2011;106(6):1081-1088.
- 65. Bajaj JS, Saeian K, Schubert CM, et al. Minimal hepatic encephalopathy is associated with motor vehicle crashes: the reality beyond the driving test. *Hepatology*. 2009;50(4):1175-1183.
- 66. Wilkinson DJ, Smeeton NJ, Watt PW. Ammonia metabolism, the brain and fatigue; revisiting the link. *Prog Neurobiol*. 2010;91(3):200-219.
- 67. Holecek M. Relation between glutamine, branched-chain amino acids, and protein metabolism. *Nutrition*. 2002;18(2):130-133.
- 68. Leweling H, Breitkreutz R, Behne F, et al. Hyperammonemia-induced depletion of glutamate and branched-chain amino acids in muscle and plasma. *J Hepatol*. 1996;25(5):756-762.
- 69. Johansen ML, Bak LK, Schousboe A, et al. The metabolic role of isoleucine in detoxification of ammonia in cultured mouse neurons and astrocytes. *Neurochem Int*. 2007;50(7-8):1042-1051.
- Rose CF. Ammonia-lowering strategies for the treatment of hepatic encephalopathy. *Clin Pharmacol Ther*. 2012;92(3):321-331.
- 71. Skowronska M, Albrecht J. Alterations of blood brain barrier function in hyperammonemia: an overview. *Neurotox Res*. 2012;21(2):236-244.
- 72. Bianchi G, Marzocchi R, Lorusso C, Ridolfi V, Marchesini G. Nutritional treatment of chronic liver failure. *Hepatol Res*. 2008;38(s1The 6 Japan Society of Hepatology Single Topic Conference: Liver Failure: Recent Progress and Pathogenesis to Management. 28-29 September 2007, Iwate, Japan):S93-S101.
- 73. Kim SJ, Kim DG, Lee MD. Effects of branched-chain amino acid infusions on liver regeneration and plasma amino acid patterns in partially hepatectomized rats. *Hepatogastroenterology*. 2011;58(109):1280-1285.
- 74. Itou M, Kawaguchi T, Taniguchi E, et al. Branched-chain amino acid supplements reduced ascites and increased the quality of life in a patient with liver cirrhosis: a case report. *Mol Med Report*. 2009;2(6):977-981.
- 75. Yamamoto M, Iwasa M, Matsumura K, et al. Improvement of regional cerebral blood flow after oral intake of branched-chain amino acids in patients with cirrhosis. *World J Gastroenterol*. 2005;11(43):6792-6799.
- 76. Iwasa M, Matsumura K, Watanabe Y, et al. Improvement of regional cerebral blood flow after treatment with branched-chain amino acid solutions in patients with cirrhosis. *Eur J Gastroenterol Hepatol*. 2003;15(7): 733-737.
- 77. Rabbani P, Prasad AS. Plasma ammonia and liver ornithine transcarbamoylase activity in zinc-deficient rats. *Am J Physiol*. 1978;235(2):E203 -E206.
- 78. Moriyama M, Matsumura H, Fukushima A, et al. Clinical significance of evaluation of serum zinc concentrations in C-viral chronic liver disease. *Dig Dis Sci*. 2006;51(11):1967-1977.
- 79. Nagasue N, Kolno H, Chang YC, Nakamura T. Iron, copper and zinc levels in serum and cirrhotic liver of patients with and without hepatocellular carcinoma. *Oncology*. 1989;46(5):293-296.
- 80. Riggio O, Merli M, Capocaccia L, et al. Zinc supplementation reduces blood ammonia and increases liver ornithine transcarbamylase activity in experimental cirrhosis. *Hepatology*. 1992;16(3):785-789.
- 81. Aquilio E, Spagnoli R, Riggio D, Seri S. Effects of zinc on hepatic ornithine transcarbamylase (OTC) activity. *J Trace Elem Electrolytes Health Dis*. 1993;7(4):240-241.
- 82. Hayashi M, Ikezawa K, Ono A, et al. Evaluation of the effects of combination therapy with branched-chain amino acid and zinc supplements on nitrogen metabolism in liver cirrhosis. *Hepatol Res*. 2007;37(8):615-619.
- 83. Takuma Y, Nouso K, Makino Y, Hayashi M, Takahashi H. Clinical trial: oral zinc in hepatic encephalopathy. *Aliment Pharmacol Ther*. 2010;32(9):1080-1090.
- 84. Ndraha S, Hasan I, Simadibrata M. The effect of L-ornithine L-aspartate and branch chain amino acids on encephalopathy and nutritional status in liver cirrhosis with malnutrition. *Acta Med Indones*. 2011;43(1):18-22.
- 85. Haussinger D. Nitrogen metabolism in liver: structural and functional organization and physiological relevance. *Biochem J*. 1990;267(2): 281-290.
- 86. Malaguarnera M, Risino C, Cammalleri L, et al. Branched chain amino acids supplemented with L-acetylcarnitine versus BCAA treatment in hepatic coma: a randomized and controlled double blind study. *Eur J Gastroenterol Hepatol*. 2009;21(7):762-770.
- 87. Matsumura T, Morinaga Y, Fujitani S, et al. Oral administration of branched-chain amino acids activates the mTOR signal in cirrhotic rat liver. *Hepatol Res*. 2005;33(1):27-32.
- 88. Kawaguchi T, Taniguchi E, Itou M, et al. The pathogenesis, complications and therapeutic strategy for hepatitis C virus-associated insulin resistance in the era of anti-viral treatment. *Rev Recent Clin Trials*. 2010;5(3): 147-157.
- 89. Kawaguchi T, Shiraishi K, Ito T, et al. Branched-chain Amino acids prolonged survival of patients with liver cirrhosis: a nationwide study in Japan [Abstract 1619]. *Hepatology*. 2012.
- Yatsuhashi H, Ohnishi Y, Nakayama S, et al. Anti-hypoalbuminemic effect of branched-chain amino acid granules in patients with liver cirrhosis is independent of dietary energy and protein intake. *Hepatol Res*. 2011;41(11):1027-1035.
- 91. Takeshita S, Ichikawa T, Nakao K, et al. A snack enriched with oral branched-chain amino acids prevents a fall in albumin in patients with liver cirrhosis undergoing chemoembolization for hepatocellular carcinoma. *Nutr Res*. 2009;29(2):89-93.
- 92. Habu D, Nishiguchi S, Nakatani S, et al. Comparison of the effect of BCAA granules on between decompensated and compensated cirrhosis. *Hepatogastroenterology*. 2009;56(96):1719-1723.
- 93. Kuwahata M, Yoshimura T, Sawai Y, et al. Localization of polypyrimidine-tract-binding protein is involved in the regulation of albumin synthesis by branched-chain amino acids in HepG2 cells. *J Nutr Biochem*. 2008;19(7):438-447.
- 94. Fukushima H, Miwa Y, Shiraki M, et al. Oral branched-chain amino acid supplementation improves the oxidized/reduced albumin ratio in patients with liver cirrhosis. *Hepatol Res*. 2007;37(9):765-770.
- 95. Togo S, Tanaka K, Morioka D, et al. Usefulness of granular BCAA after hepatectomy for liver cancer complicated with liver cirrhosis. *Nutrition*. 2005;21(4):480-486.
- 96. Sato S, Watanabe A, Muto Y, et al. Clinical comparison of branched-chain amino acid (l-Leucine, l-Isoleucine, l-Valine) granules and oral nutrition for hepatic insufficiency in patients with decompensated liver cirrhosis (LIV-EN study). *Hepatol Res*. 2005;31(4):232-240.
- 97. Nishitani S, Takehana K. Pharmacological activities of branched-chain amino acids: augmentation of albumin synthesis in liver and improvement of glucose metabolism in skeletal muscle. *Hepatol Res*. 2004;30S:19-24.
- 98. Nishiguchi S, Habu D. Effect of oral supplementation with branchedchain amino acid granules in the early stage of cirrhosis. *Hepatol Res*. 2004;30S:36-41.
- 99. Kuwahata M, Kuramoto Y, Tomoe Y, et al. Posttranscriptional regulation of albumin gene expression by branched-chain amino acids in rats with acute liver injury. *Biochim Biophys Acta*. 2004;1739(1):62-69.
- 100. Ijichi C, Matsumura T, Tsuji T, Eto Y. Branched-chain amino acids promote albumin synthesis in rat primary hepatocytes through the mTOR signal transduction system. *Biochem Biophys Res Commun*. 2003;303(1):59-64.
- 101. Habu D, Nishiguchi S, Nakatani S, et al. Effect of oral supplementation with branched-chain amino acid granules on serum albumin level in the early stage of cirrhosis: a randomized pilot trial. *Hepatol Res*. 2003;25(3):312-318.
- 102. Fukushima H, Miwa Y, Ida E, et al. Nocturnal branched-chain amino acid administration improves protein metabolism in patients with liver cirrhosis: comparison with daytime administration. *JPEN J Parenter Enteral Nutr*. 2003;27(5):315-322.
- 103. Tayek JA, Bistrian BR, Hehir DJ, et al. Improved protein kinetics and albumin synthesis by branched chain amino acid-enriched total parenteral nutrition in cancer cachexia. A prospective randomized crossover trial. *Cancer*. 1986;58(1):147-157.
- 104. Sugiyama K, Yu L, Nagasue N. Direct effect of branched-chain amino acids on the growth and metabolism of cultured human hepatocellular carcinoma cells. *Nutr Cancer*. 1998;31(1):62-68.
- 105. Muto Y, Sato S, Watanabe A, et al. Overweight and obesity increase the risk for liver cancer in patients with liver cirrhosis and long-term oral supplementation with branched-chain amino acid granules inhibits liver carcinogenesis in heavier patients with liver cirrhosis. *Hepatol Res*. 2006;35(3):204-214.
- 106. Kobayashi M, Ikeda K, Arase Y, et al. Inhibitory effect of branched-chain amino acid granules on progression of compensated liver cirrhosis due to hepatitis C virus. *J Gastroenterol*. 2008;43(1):63-70.
- 107. Yoshiji H, Noguchi R, Kitade M, et al. Branched-chain amino acids suppress insulin-resistance-based hepatocarcinogenesis in obese diabetic rats. *J Gastroenterol*. 2009;44(5):483-491.
- 108. Harima Y, Yamasaki T, Hamabe S, et al. Effect of a late evening snack using branched-chain amino acid-enriched nutrients in patients undergoing hepatic arterial infusion chemotherapy for advanced hepatocellular carcinoma. *Hepatol Res*. 2010;40(6):574-584.
- 109. Iwasa J, Shimizu M, Shiraki M, et al. Dietary supplementation with branched-chain amino acids suppresses diethylnitrosamine-induced liver tumorigenesis in obese and diabetic C57BL/KsJ-db/db mice. *Cancer Sci*. 2010;101(2):460-467.
- 110. Kuroda H, Ushio A, Miyamoto Y, et al. Effects of branched-chain amino acid-enriched nutrient for patients with hepatocellular carcinoma following radiofrequency ablation: a one-year prospective trial. *J Gastroenterol Hepatol*. 2010;25(9):1550-1555.
- 111. Yoshiji H, Noguchi R, Kaji K, et al. Attenuation of insulin-resistancebased hepatocarcinogenesis and angiogenesis by combined treatment with branched-chain amino acids and angiotensin-converting enzyme inhibitor in obese diabetic rats. *J Gastroenterol*. 2010;45(4):443-450.
- 112. Hayaishi S, Chung H, Kudo M, et al. Oral branched-chain amino acid granules reduce the incidence of hepatocellular carcinoma and improve eventfree survival in patients with liver cirrhosis. *Dig Dis*. 2011;29(3):326-332.
- 113. Lee IJ, Seong J, Bae JI, et al. Effect of oral supplementation with branched-chain amino acid (BCAA) during radiotherapy in patients with hepatocellular carcinoma: a double-blind randomized study. *Cancer Res Treat*. 2011;43(1):24-31.
- 114. Ninomiya S, Shimizu M, Imai K, et al. Possible role of visfatin in hepatoma progression and the effects of branched-chain amino acids on visfatin-induced proliferation in human hepatoma cells. *Cancer Prev Res (Phila)*. 2011;4(12):2092-2100.
- 115. Yoshiji H, Noguchi R, Ikenaka Y, et al. Combination of branched-chain amino acids and angiotensin-converting enzyme inhibitor suppresses the cumulative recurrence of hepatocellular carcinoma: a randomized control trial. *Oncol Rep*. 2011;26(6):1547-1553.
- 116. Hagiwara A, Nishiyama M, Ishizaki S. Branched-chain amino acids prevent insulin-induced hepatic tumor cell proliferation by inducing apoptosis through mTORC1 and mTORC2-dependent mechanisms. *J Cell Physiol*. 2012;227(5):2097-2105.
- 117. Ichikawa K, Okabayashi T, Maeda H, et al. Oral supplementation of branched-chain amino acids reduces early recurrence after hepatic resection in patients with hepatocellular carcinoma: a prospective study. *Surg Today*. 2013;43:720-726.
- 118. Morihara D, Iwata K, Hanano T, et al. Late-evening snack with branchedchain amino acids improves liver function after radiofrequency ablation for hepatocellular carcinoma. *Hepatol Res*. 2012;42(7):658-667.
- 119. Nishikawa H, Osaki Y, Inuzuka T, et al. Branched-chain amino acid treatment before transcatheter arterial chemoembolization for hepatocellular carcinoma. *World J Gastroenterol*. 2012;18(12):1379-1384.
- 120. Okamoto M, Sakaida I, Tsuchiya M, Suzuki C, Okita K. Effect of a late evening snack on the blood glucose level and energy metabolism in patients with liver cirrhosis. *Hepatol Res*. 2003;27(1):45-50.
- 121. Sakaida I, Tsuchiya M, Okamoto M, Okita K. Late evening snack and the change of blood glucose level in patients with liver cirrhosis. *Hepatol Res*. 2004;30S:67-72.
- 122. Tsuchiya M, Sakaida I, Okamoto M, Okita K. The effect of a late evening snack in patients with liver cirrhosis. *Hepatol Res*. 2005;31(2):95-103.
- 123. Kawaguchi T, Taniguchi E, Itou M, et al. Branched-chain amino acids improve insulin resistance in patients with hepatitis C virus-related liver disease: report of two cases. *Liver Int*. 2007;27(9):1287-1292.
- 124. Urata Y, Okita K, Korenaga K, et al. The effect of supplementation with branched-chain amino acids in patients with liver cirrhosis. *Hepatol Res*. 2007;37(7):510-516.
- 125. Korenaga K, Korenaga M, Uchida K, Yamasaki T, Sakaida I. Effects of a late evening snack combined with alpha-glucosidase inhibitor on liver cirrhosis. *Hepatol Res*. 2008;38(11):1087-1097.
- 126. Kawaguchi T, Yamagishi S, Sata M. Branched-chain amino acids and pigment epithelium-derived factor: novel therapeutic agents for hepatitis c virus-associated insulin resistance. *Curr Med Chem*. 2009;16(36):4843- 4857.
- 127. Higuchi N, Kato M, Miyazaki M, et al. Potential role of branched-chain amino acids in glucose metabolism through the accelerated induction of the glucose-sensing apparatus in the liver. *J Cell Biochem*. 2011;112(1):30-38.
- 128. Miyake T, Abe M, Furukawa S, et al. Long-term Branched-chain amino acid supplementation improves glucose tolerance in patients with nonalcoholic steatohepatitis-related cirrhosis. *Intern Med*. 2012;51(16):2151- 2155.
- 129. Takegoshi K, Nanasawa H, Itoh H, et al. Effects of branched-chain amino acid-enriched nutrient mixture on natural killer cell activity in viral cirrhosis. *Arzneimittelforschung*. 1998;48(6):701-706.
- 130. Nakamura I, Ochiai K, Imawari M. Phagocytic function of neutrophils of patients with decompensated liver cirrhosis is restored by oral supplementation of branched-chain amino acids. *Hepatol Res*. 2004;29(4):207-211.
- 131. Taniguchi E, Kawaguchi T, Shimada M, et al. Branched-chain amino acid supplementation complements conventional treatment for spontaneous bacterial peritonitis. *Dig Dis Sci*. 2006;51(6):1057-1060.
- 132. Kakazu E, Kanno N, Ueno Y, Shimosegawa T. Extracellular branchedchain amino acids, especially valine, regulate maturation and function of monocyte-derived dendritic cells. *J Immunol*. 2007;179(10):7137-7146.
- 133. Nakamura I, Ochiai K, Imai Y, Moriyasu F, Imawari M. Restoration of innate host defense responses by oral supplementation of branchedchain amino acids in decompensated cirrhotic patients. *Hepatol Res*. 2007;37(12):1062-1067.
- 134. Itou M, Kawaguchi T, Taniguchi E, et al. Heating improves poor compliance with branched chain amino acid-rich supplementation in patients with liver cirrhosis: a before-after pilot study. *Mol Med Report*. 2009;2(6):983-987.
- 135. Kakazu E, Ueno Y, Kondo Y, et al. Branched chain amino acids enhance the maturation and function of myeloid dendritic cells ex vivo in patients with advanced cirrhosis. *Hepatology*. 2009;50(6):1936-1945.
- 136. Tomiya T, Omata M, Fujiwara K. Branched-chain amino acids, hepatocyte growth factor and protein production in the liver. *Hepatol Res*. 2004;30S:14-18.
- 137. Tomiya T, Omata M, Fujiwara K. Significance of branched chain amino acids as possible stimulators of hepatocyte growth factor. *Biochem Biophys Res Commun*. 2004;313(2):411-416.
- 138. Okabayashi T, Nishimori I, Sugimoto T, et al. Effects of branchedchain amino acids-enriched nutrient support for patients undergoing liver resection for hepatocellular carcinoma. *J Gastroenterol Hepatol*. 2008;23(12):1869-1873.
- 139. Koreeda C, Seki T, Okazaki K, Ha-Kawa S K, Sawada S. Effects of late evening snack including branched-chain amino acid on the function of hepatic parenchymal cells in patients with liver cirrhosis. *Hepatol Res*. 2011;41(5):417-422.
- 140. Kuwahata M, Kubota H, Kanouchi H, et al. Supplementation with branched-chain amino acids attenuates hepatic apoptosis in rats with chronic liver disease. *Nutr Res*. 2012;32(7):522-529.
- 141. Miuma S, Ichikawa T, Arima K, et al. Branched-chain amino acid deficiency stabilizes insulin-induced vascular endothelial growth factor mRNA in hepatocellular carcinoma cells. *J Cell Biochem*. 2012;113(10):3113- 3121.
- 142. Nakanishi C, Doi H, Katsura K, Satomi S. Treatment with L-valine ameliorates liver fibrosis and restores thrombopoiesis in rats exposed to carbon tetrachloride. *Tohoku J Exp Med*. 2010;221(2):151-159.
- 143. Yoshiji H, Noguchi R, Ikenaka Y, et al. Combination of branched-chain amino acid and angiotensin-converting enzyme inhibitor improves liver fibrosis progression in patients with cirrhosis. *Mol Med Report*. 2012;5(2):539-544.
- 144. Ohno T, Tanaka Y, Sugauchi F, et al. Suppressive effect of oral administration of branched-chain amino acid granules on oxidative stress and inflammation in HCV-positive patients with liver cirrhosis. *Hepatol Res*. 2008;38(7):683-688.
- 145. Kuwahata M, Kubota H, Katsukawa M, et al. Effect of branched-chain amino acid supplementation on the oxidized/reduced state of plasma albumin in rats with chronic liver disease. *J Clin Biochem Nutr*. 2012;50(1):67-71.
- 146. Honda M, Takehana K, Sakai A, et al. Malnutrition impairs interferon signaling through mTOR and FoxO pathways in patients with chronic hepatitis C. *Gastroenterology*. 2011;141(1):128-140.
- 147. Kawaguchi T, Torimura T, Takata A, Satomi S, Sata M. Valine, a branched-chain amino acid, reduced HCV viral load and led to eradication of HCV by interferon therapy in a decompensated cirrhotic patient: a case report. *Case Rep Gastroenterol*. 2012;6(3):660-667.
- 148. Kawamura N, Nakajima H, Takashi SI. Administration of granulated BCAA and quality of life. *Hepatol Res*. 2004;30(S):42-45.
- 149. Nakaya Y, Okita K, Suzuki K, et al. BCAA-enriched snack improves nutritional state of cirrhosis. *Nutrition*. 2007;23(2):113-120.
- 150. Ichikawa T, Naota T, Miyaaki H, et al. Effect of an oral branched chain amino acid-enriched snack in cirrhotic patients with sleep disturbance. *Hepatol Res*. 2010;40(10):971-978.
- 151. Okabayashi T, Iyoki M, Sugimoto T, Kobayashi M, Hanazaki K. Oral supplementation with carbohydrate- and branched-chain amino acid-enriched nutrients improves postoperative quality of life in patients undergoing hepatic resection. *Amino Acids*. 2011;40(4):1213-1220.
- 152. Hidaka H, Nakazawa T, Kutsukake S, et al. The efficacy of nocturnal administration of branched-chain amino acid granules to improve quality of life in patients with cirrhosis. *J Gastroenterol*. 2013;48(2):269-276.